

pared to amylose. The hydrolysis of amylopectin and amylose is compared in Fig. 1, which graphically shows that the hydrolysis rate for the amylopectin decreased abruptly to a definitely lower value as the level of 60% conversion was approached, a striking analogy to the well known behavior of β -amylase on this substrate. These results may be taken as further evidence that the amylo-glucosidase does not operate at random but, rather, by a terminalwise attack on the non-reducing end of starch molecules, and that the rate of hydrolysis for anomalous linkages, at points of branching, is very much less than the rate for normal glucoside linkages.

It appears quite possible in light of these experiments, that the limit-dextrinase activity noted in early work for some relatively crude enzyme preparations may have been due in part at least to the combined action of an amylo-glucosidase and an α -amylase.

The very large and no doubt slightly branched amyloses, such as tapioca A-fraction, gave hydrolysis rates intermediate between that of corn amylose and amylopectin. Using *equimolar* concentrations, the rate of hydrolysis for these larger amylose molecules, as mg. of glucose produced per unit time, was initially greater than the rate for corn amylose, but decreased to values approaching that of the corn samples as hydrolysis progressed. These experiments will be detailed in a future communication.²²

It is possible that the final amylo-glucosidase preparation, which was used to obtain the data in Tables VI and VII and for Fig. 1, still contained a

(22) R. W. Kerr and F. C. Cleveland, manuscript in preparation.

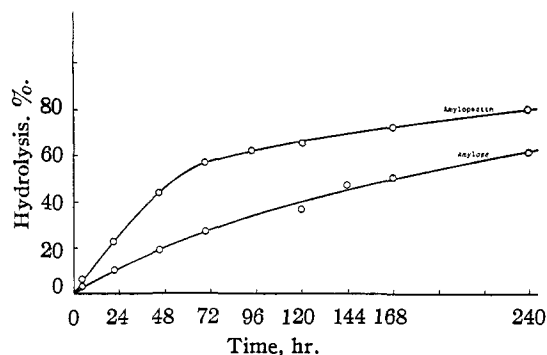


Fig. 1.—Hydrolyses of amylose and amylopectin at equal weight concentration by amylo-glucosidase.

trace of α -amylase activity. Even a trace of α -amylase would modify the results and may account for the slight drop in peak light absorption values during the corn A-fraction hydrolysis as well as a part, at least, of the secondary phase of the amylopectin hydrolysis. However, it is believed that the data given in Tables VI and VII, when considered together with the data in preceding tables, showing the manner in which the kinetics of the hydrolysis tend to change as the enzyme preparation was progressively freed of α -amylase activity and the residual activity became more exclusively that of the amylo-glucosidase factor, clearly indicate the general pattern of the action of this latter enzyme and distinguish it from other carbohydrases.

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The Synthesis of C¹⁴-Labeled Glycerol^{1,2}

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Glycerol-1,3-C₂¹⁴ has been prepared in over-all yield of 19%, by condensing nitromethane with paraformaldehyde-C¹⁴, followed by reduction and diazotization.

Glycerol, labeled with carbon-14, was needed for projected studies in this Laboratory on the metabolism of glycerides. Although there are several synthetic routes³ which might have been employed, the scheme shown in Fig. 1 was chosen because of the availability of both nitromethane-C¹⁴⁴ and formaldehyde-C¹⁴.⁵

Experimental

Sodium-2-nitro-1,3-propanediol (I).—According to the procedure of Schmidt and Wilkendorf,⁶ 13.47 g. of paraformaldehyde-C¹⁴ (0.449 mole)⁷ was allowed to react with

(1) This investigation was supported in part by a grant from the United States Atomic Energy Commission.

(2) Presented at the 118th Meeting of the American Chemical Society, Chicago, Illinois, September, 1950.

(3) Since the writing of this manuscript the preparation of glycerol-1-C¹⁴ was reported with a yield in radioactivity of 12.3%: A. P. Doerschuk, *THIS JOURNAL*, **73**, 821 (1951).

(4) L. G. Sowden, *J. Biol. Chem.*, **180**, 56 (1949).

(5) A. R. Jones and W. J. Skraba, *Science*, **110**, 332 (1949).

(6) E. Schmidt and R. Wilkendorf, *Ber.*, **52**, 395 (1919).

(7) Isotopes Division, U. S. Atomic Energy Commission, Oak Ridge, Tenn. The material was lumpy and sticky, whereas Eastman paraformaldehyde is powdery. Its activity was specified as approximately 5 microcuries per millimole.

8.54 g. of nitromethane (0.14 mole) in 110 ml. of absolute methanol by the addition of 8–10 drops of 50% KOH. When Eastman Kodak Co. paraformaldehyde was used, the reaction began immediately upon heating on a steam-cone, and with mechanical stirring, the solution became clear within ten minutes. When the reaction was performed with the isotopic paraformaldehyde, there were still some unreacted particles after 45 minutes heating. After decanting from the insoluble particles the mixture was cooled to -5° . A solution containing 4.02 g. of sodium dissolved in 65 ml. of absolute methanol was added during a period of 20 minutes with mechanical stirring. After standing for six hours at -5° , the precipitate was filtered off. The yield of the sodium salt of 2-nitro-1,3-propanediol, containing 2 molecules of methanol of crystallization, was 27 g. or 58%. The filtrate was neutralized with 7.8 N HCl in methanol, mixed with the unreacted paraformaldehyde and the whole procedure was repeated by adding 3.05 g. (0.05 mole) of nitromethane. The yield in this recycling was 8 g. A second recycling yielded 1 g. The total yield was 36 g. or 77% based on the paraformaldehyde.

2-Amino-1,3-propanediol Hydrochloride (II).—Hydrogenation of I was carried out in two batches of about 18 g. each, using an apparatus similar to the Parr apparatus. Each batch was dissolved as completely as possible in 225 ml. of absolute methanol, and 2.1 equivalents of glacial acetic acid was added, followed by 4 g. of Raney nickel. The pressure was kept between 37 and 20 lb./sq. in. Hy.

